### Simon Dedman

### gbm.auto

Automated Boosted Regression Tree software for spatial prediction of multiple life-history stock components

simondedman@gmail.com
simondedman.com

### Background / specific case for work

- Spatial approaches to manage data-poor species
- Existing techniques often struggle / suboptimal
- Boosted Regression Trees (BRTs/GBMs) complicated but excellent performance:
  - Robust to poor/absent data
  - Can use abundance data
  - Unaffected by missing predictor values, outliers, multicollinearity
  - Can accommodate large numbers of explanatory variables without penalty
  - More robust predictions than GLMs and GAMs
  - Less variance (oversensitivity to noise leading to overfitting/imprecision)
  - Less bias (false assumptions in the algorithm leading to underfitting/inaccuracy)
  - Lower risk of misspecification
  - Ability to model complex interactions

#### Regression Tree models:

- Machine learning. No assumed relationship, model learns predictor-response relationships
- Uses algorithms to partition the predictor space into sections of the most homogenous response to predictors blocks of reliable predictor-response relationship carving out these blocks in binary splits at points along the predictors' ranges
- Predictors & split points calculated to minimise prediction error
- Not as accurate as GLMs/GAMs
- Bad at modelling smooth functions
- Very dependent on the sample data used, i.e. results aren't generalisable

### **Boosting**

- "it is easier to find and average many rough rules of thumb, than to find a single, highly accurate prediction rule" (Elith et al. 2008)
- Finds one tree that best explains the predictors-response relationship, then
- Finds the tree that best explains the predictors-response relationship of the residuals of the one-tree model (which is a new tree with different values)
- Updates the model to incorporate the predictors-response relationship information gained from tree 1 plus tree 2
- Runs this new 2 tree model on the data (choosing a different random testing chunk each time), producing new residuals. Makes new tree to test residuals, adds to model to make 3 tree model, runs 3 tree model. Repeats 1000s of times: remaining unaccounted-for deviance falls, until adding trees is unhelpful.

### **Journal of Animal Ecology**

British Ecological Society

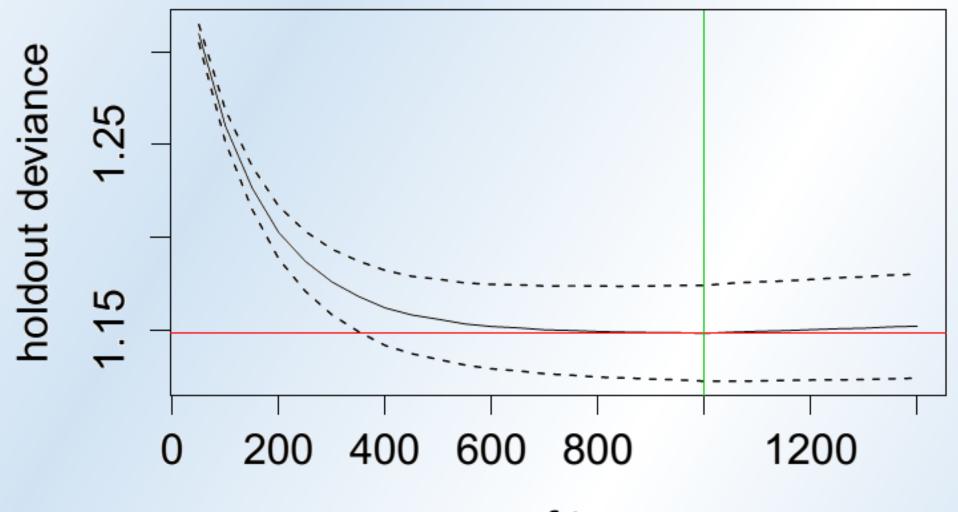
Journal of Animal Ecology 2008, 77, 802-813

#### doi: 10.1111/j.1365-2656.2008.01390.x

#### A working guide to boosted regression trees Trees 1 & 2 Tree 1 J. Elith<sup>1</sup>\*, J. R. Leathwick<sup>2</sup> and T. Hastie<sup>3</sup> Marginal effect on logit(p) Marginal effect on logit(p) $X_{1} \leq t_{1}$ $X_2 \leq t_2$ $X_1 \ge t_2$ 0 $X_2 \ge t_4$ 14 18 10 14 18 10 Summer temp (°C) Summer temp (°C) Trees 1 to 1050 Trees 1 to 100 Marginal effect on logit(p) Marginal effect on logit(p) 0 0 10 14 18 18 10 14 Summer temp (°C) Summer temp (°C)

"Boosting is a numerical optimization technique for minimizing the loss function by adding, at each step, a new tree that best reduces (steps down the gradient of) the loss function. For BRT, the first regression tree is the one that, for the selected tree size, maximally reduces the loss function. For each following step, the focus is on the residuals: at the second step, a tree is fitted to the residuals of the first tree, and that second tree could contain quite different variables and split points compared with the first. The model is then updated to contain two trees (two terms), and the residuals from this two-term model are calculated, and so on."

As the model incorporates more trees, the remaining unaccounted-for deviance falls, until the point where adding more trees adds unnecessary complexity and explains the predictors-response relationship LESS well. The code notes the number of trees which produce the lowest holdout deviance score, here 1000, and uses that model going forward.

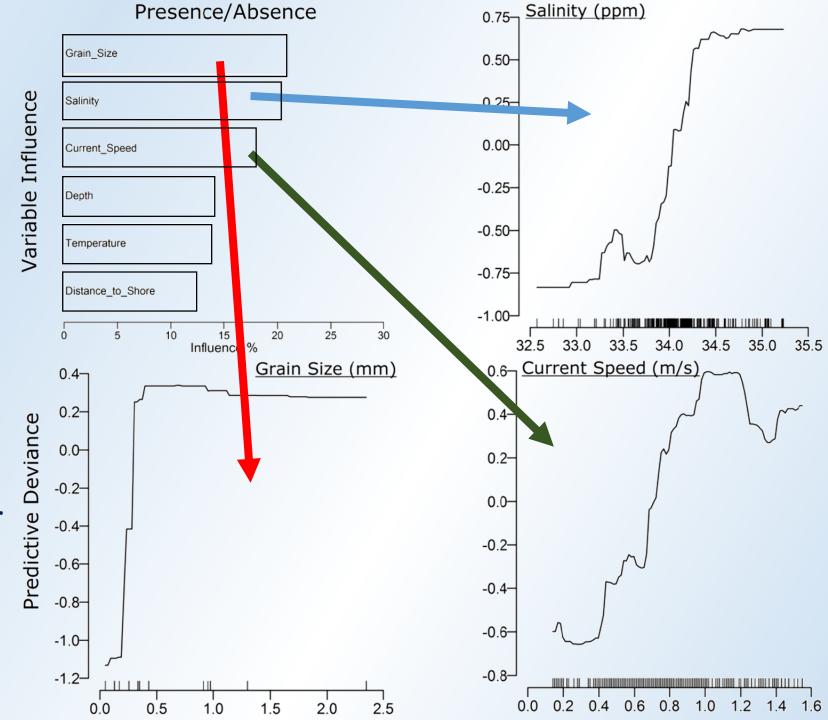


no. of trees

Relative contribution of each variable

Predictor-response relationship for each variable

All of this complex information lives within the built model object. It's not completely a black box: you CAN force it to divulge its secrets, such as these figures, but its real value is using its knowledge to make predictions.



# Work done to address specific need

- Software suite in R that automates and greatly simplifies delta log-normal Boosted Regression Tree spatial modelling.
- Powerful statistical modelling technique made accessible to potential users in the ecological and modelling communities.
- Input data: Ray survey CPUE disaggregated into: Positive data only, Presence/ log-normalised Absence Input data: Environmental covariates (Depth, distance to shore, temperature, salinity, current speed, substrate grain size) **Iterative parameter** BRT model 2 BRT model 1 optimisation and model simplification **Output graphs:** Distance to shore Relative Current speed Temperature influence Salinity Best abundance Best presence/ Depth δ 5 10 15 20 25 30 Influence % absence model model, log reversed Salinity Mr Predictive deviance Presence probability x abundance = **Output maps:** Whole area CPUE prediction **Predicted abundance** Representativeness

		Import data, specify predictor & response vo Set process control & design variables (optic	
	gbm.utils 🔸	<ul> <li>Checks and acquires packages</li> </ul>	
		Zero-inflated data check Pre-processes data (binary & log-normal pre	sence-only)
	gbm.auto	Names & models current variable combo	
	dismo (+)	Continuously selects best model	
		Simplifies model —	→ gbm.simplify
	gbm	Tests simplification	
	0	Outputs line plots (together/separate)	- gbm.plot
		Outputs dot plots	nlot abm
gb	m.map	Outputs 3D plots [pending]	<ul> <li>plot.gbm</li> </ul>
gbm.predict.grids		Outputs relative influence bar plots Outputs relative influence CSV file	gbm.plot.fits
		Outputs prediction map	gbm.perspec
gbm.rsb		Outputs representativeness map	gumperspec
gbm.map		Outputs CSV file of prediction data	
		) Outputs report	-gbm.interactions
		Loops the next variable combination	

## Acquiring global coastlines with gbm.basemap

mybounds <- c(range(samples[,3]),range(samples[,2]))
gbm.basemap(bounds = mybounds)</pre>

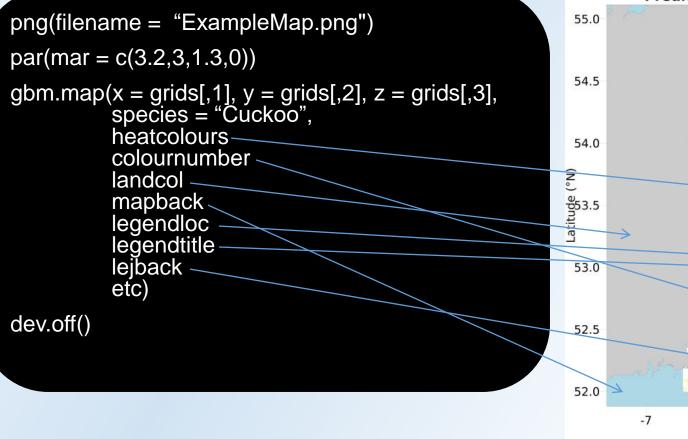
Resolution 1, "coarse" Resolution 5, "full"

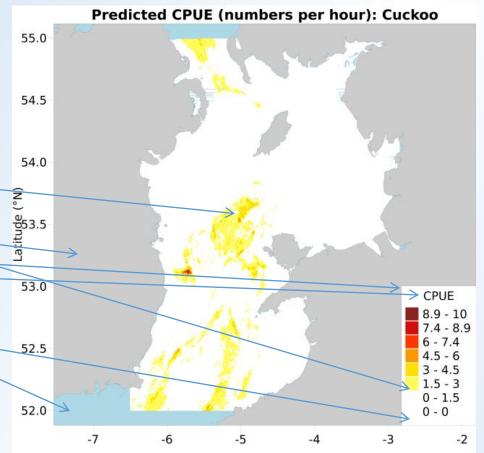
GBM = Gradient Boosting Machine / Generalised Boosted Models: EXACTLY the same thing as BRT Boosted Regressions Trees, but a different name. And all the parameters have different name. No idea why this issue exists.

	А	В	С
1	LONGITUDE	LATITUDE	Abundance?
2	-6.45765	53.97035	0
3	-6.18835	52.2358	1
4	-6.1692	52.57755	0
5	-6.11505	52.7634	0
6	-6.115	52.7534	2
7	-6.1147	52.7636	3
8	-6.11315	53.6292	2
9	-6.1128	53.62605	0
10	-6.1128	54.0279	0
11	-6.11245	52.75085	2
12	-6.11245	52.7559	0

## Mapping with gbm.map

- Mapping function for gridded data
- calculates the cell size automatically
- allows user to alter most elements of the output





Longitude (°W)

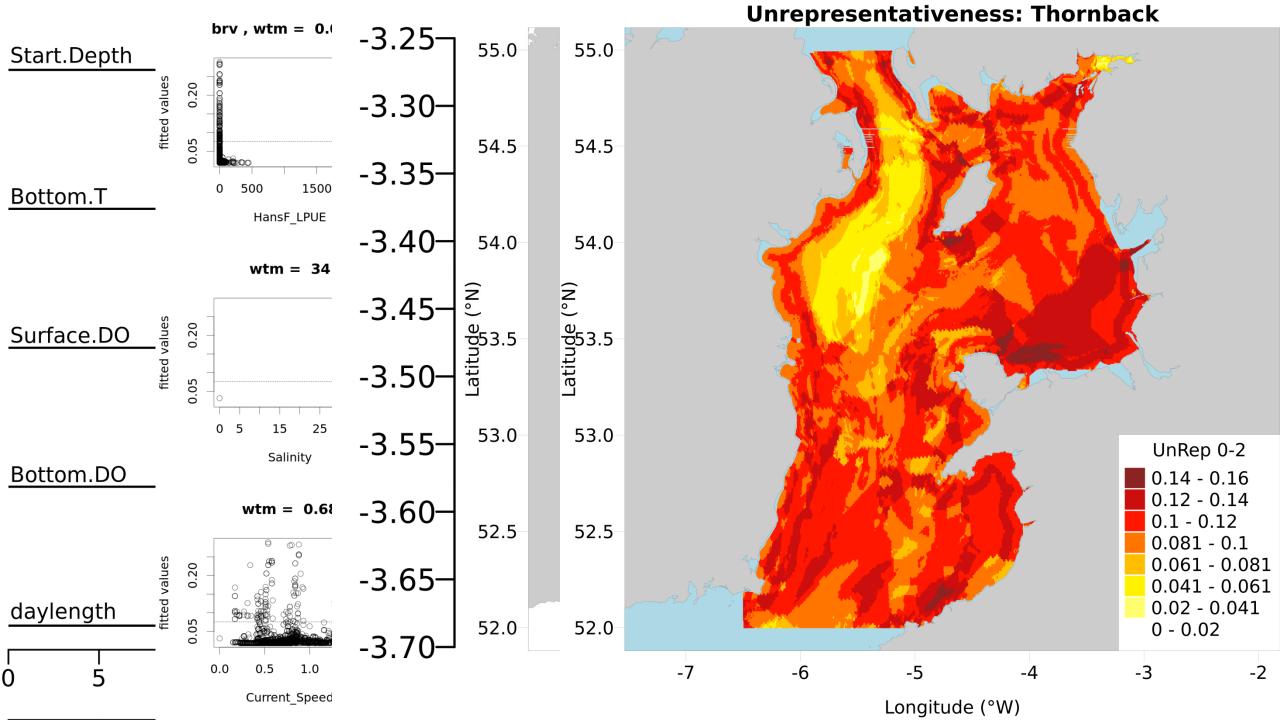
### Abundance predictions with gbm.auto

samples <- read.csv("samples.csv")</pre>

```
grids <- read.csv("grids.csv")
```

```
gbm.auto(samples = samples, grids = grids, expvar = 4:6, resvar = 3)
```

- Uses gbm.basemap, gbm.map, gbm.rsb and various other functions
- Allows the user to specify which data distribution to use
- can check for zero-inflation and transform data to use the deltalognormal model on long-tailed zero-inflated data
- automatically loops through the user-set combinations of parameters and multiple response variables



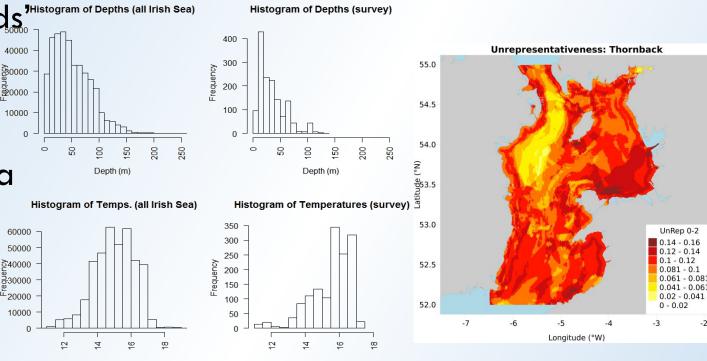
Explanatory Variables		Response Va	ariables Zero Inflated?		Bin_BRT.tc2.lr0.001.bf0.5		Best Binary BRT	
Year F.blacknose			TRUE	trees: 1400		Model combo: Bin_BRT.tc2.lr0.001.bf0.5		
Season					Training Data Correlation: 0.588134932326192		Model CV score: 0.588134932326192	
Lon					CV Mean Deviance: 1.08280416606036		Training data AUC score: 0.8827	
Surface.T					CV Deviance SE: 0.0478621899131645		CV AUC score: 0.6778	
Bottom.T					CV Mean Correlation: 0.27417021183714		CV AUC se: 0.0591583505892824	
Surface.DO					CV Correlation SE: 0.0886817493754063			
Bottom.DO								
Start.Depth								
Bin_BRT_simp predictors kept (ordered)			Bin_BR1	_simp predictors dropped	Simplified I	plified Binary BRT stats		
Season				Surface.DO tr		trees: 2600		
Depth.Bin				Start.Depth		Training Data Correlation: 0.564892280252173		
Year					CV Mean D		eviance: 1.04457745738497	
Surface.T					CV Deviand		e SE: 0.0497216516054207	
Bottom.T					CV Mean Correlation: 0.305782672406723			
uavielieur	ength Best Binary BRT variables Relative Influ				-CV Correlation SE: 0.0775726580103382			
	Start.Depth	1			Start.Depth and Lon. Size: 1.18	-		
	ottom.DO Surface.DO			5.41003404	503265 Bottom.DO and Surface.DO. Size: 0.25			
2011	Lon				The area under the ROC curve can be integrated and interpreted as an Area Under			
	7 0		0.070503635 7.959983717	the Curve (AUC) value that has a range from 0.5 to 1. Using this metric, a value				
	Bottom.T			7.63123564	<ul> <li>one indicates perfect discrimination of probabilities between presence and abser</li> <li>samples and a value of 0.5 indicates that model discrimination is no better than</li> </ul>		abilities between presence and absence	
	Surface.T			3.550914543			odel discrimination is no better than	
	Year		2.092		random. While models with AUC values greater than 0.6 are considered useful			
	Depth.Bin		1	.417916858	(Parisien and Moritz 2009), values greater than 0.8 are considered very good, and above greater than 0.9 excellent (Lane et al. 2009).			
	Season			.427653375				

# Visual assessment of data quality and representativeness with *gbm.rsb*

Temperature (°C

**Representativeness Surface Builder** 

- Compares frequency distribution of the explanatory variables from the 'grids' History data with those from the 'samples'
- Differences summed into a score indicating how well the samples data captures that variable's full range
- Calculated for every cell in 'grids'
- Exported to csv & mapped with gbm.map
- Higher values = poor coverage = be more cautious with conclusions at that point.

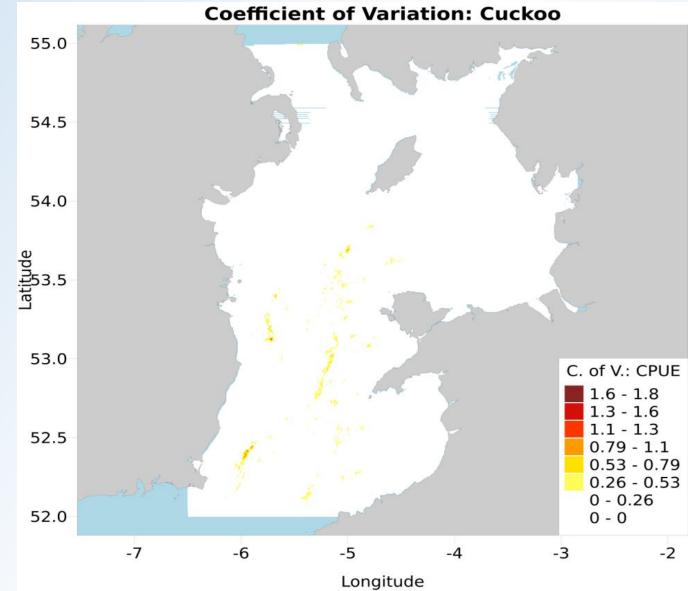


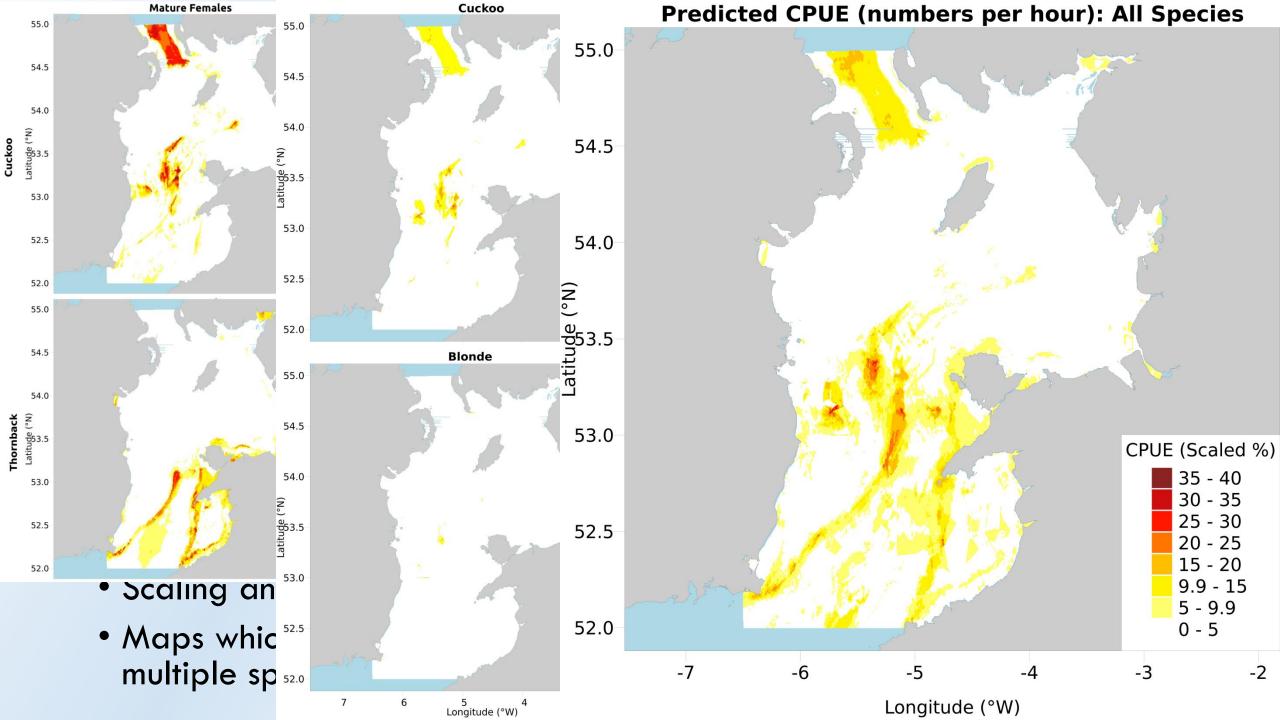
Temperature (°C)

gbm.rsb(samples, grids, expvarnames, gridslat, gridslon)

# Calculating the coefficient of variation of predicted abundance with *gbm.loop*

- Repeats gbm.auto run a userspecified number of times
- Calculates and plots the minimum, average, maximum, and variance of the variable influence values (bar plot data)
- Calculates and plots the minimum, average, and maximum partial dependence values (line plot data)
- Calculates coefficient of variation for predicted abundance map.
- Produces map and csv files

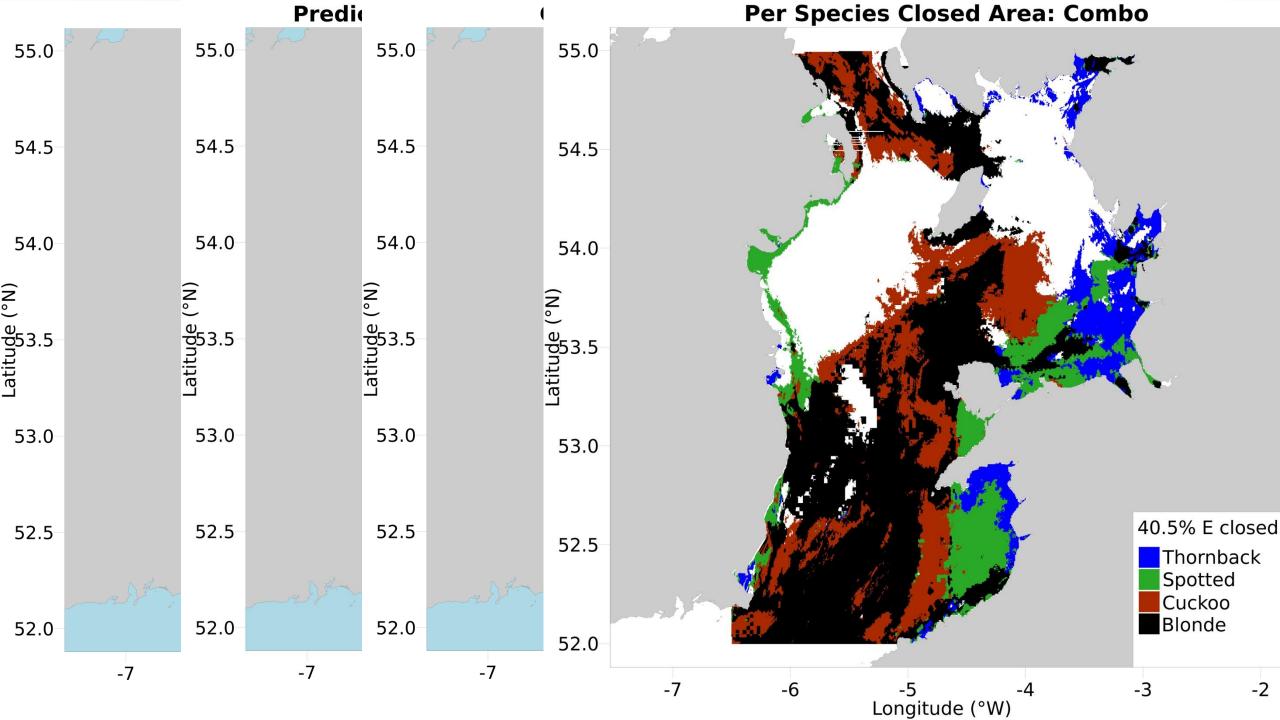




### Generating MPAs with gbm.valuemap

gbm.valuemap(dbase = mydata, goodcols = c(5,3,6,4), badcols = 7, conservecol = 8, HRMSY = c(0.14,0.08,0.08,0.15))
Predictive maps only addresses half the problem.

- Conservation plans are prioritisations: must consider socioeconomic metrics e.g. fishing effort
- Need biologically-derived MPA candidates. Maximum Sustainable Yield (MSY) principle of escapement biomass: percentage to retain annually to conserve the stock, Harvest Rate at MSY ( $HR_{MSY}$ ).
- Predicted abundance map of rays Vs map of fishing effort = areas to preferentially conserve, and areas to avoid closing to minimise effort displacement.
- Cumulatively add cells sorted from most to least preferable to close until you have an MPA big enough to protect the most conservationally valuable species' HR<sub>MSY</sub> ("species 1")
- Do the same with Species 2, but with Species 1's MPA already in place, i.e. you just grow Species 1's MPA until it protects Species 2. Repeat for all species.
- Instead of 'abundance Vs effort' prioritisation sort, can sort by effort only, abundance only, or conservation map areas from gbm.cons



### Pre-run parameter scoping with gbm.bfcheck

gbm.bfcheck(samples = samples, resvar = 3)

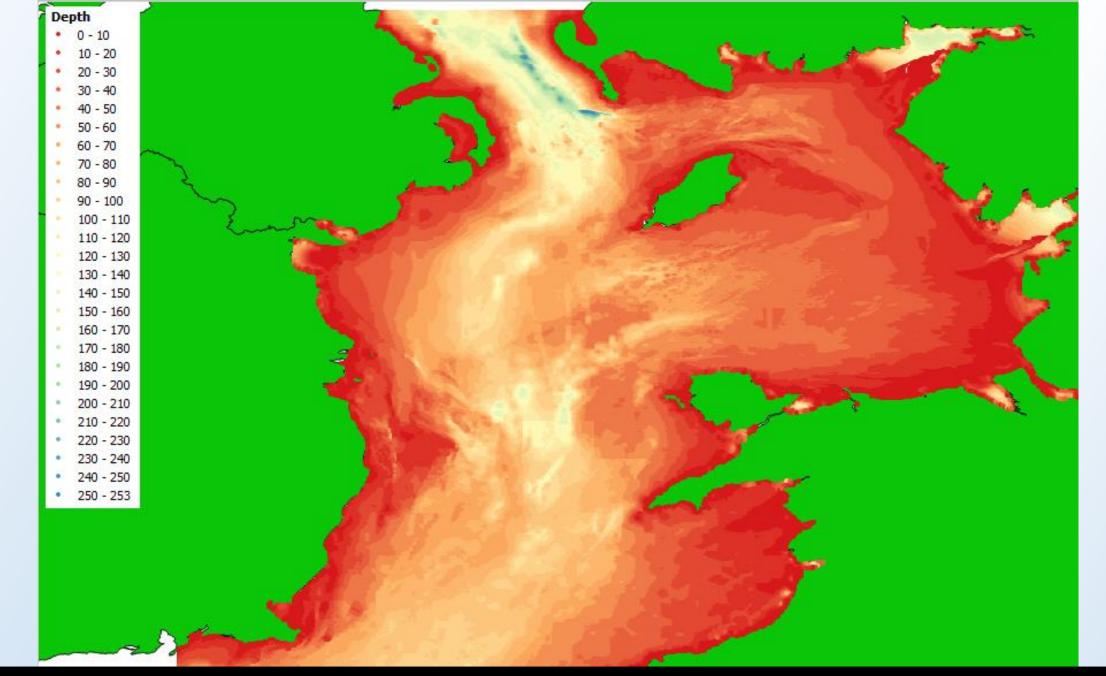
- Calculates the minimum binary and Gaussian BRT bag fraction sizes
- Users can check and optimise BFs before starting gbm.auto runs

### Conclusions

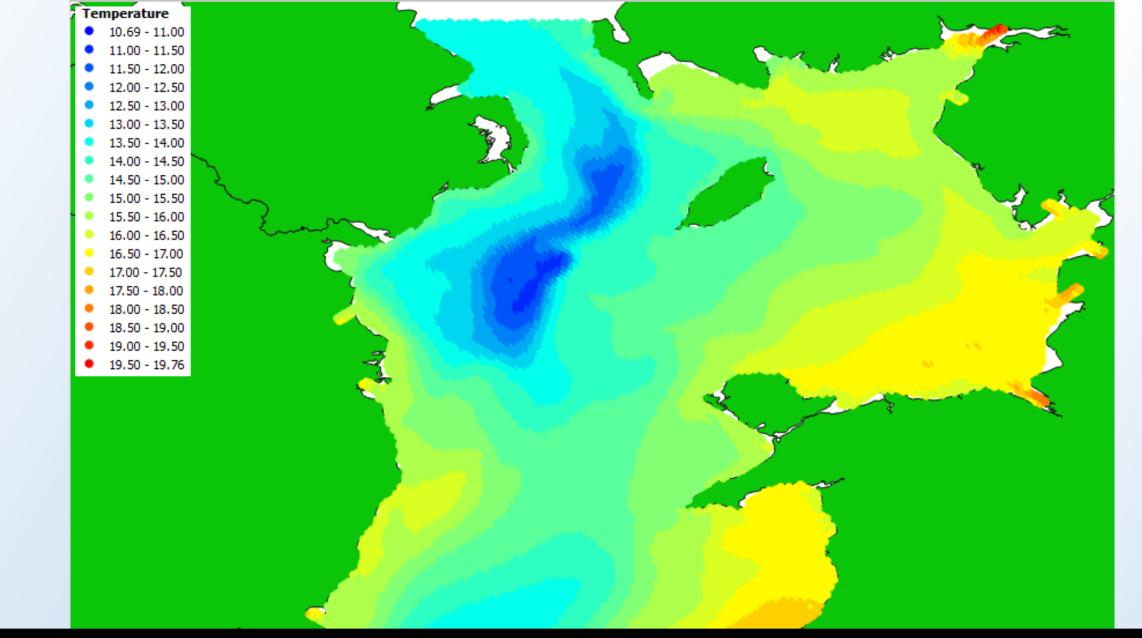
- Easily-usable and feature-rich resource.
- Data-poor or rich; single or multiple species or subsets.
- Users can easily produce predicted abundance maps, explanatory variable diagnoses, conservation priority area maps and area closure proposals, with little work or prior knowledge required.
- Facilitate and expedite conservation of data-poor species using MPAs that balance competing priorities with the full engagement of stakeholders.
- Customisability means users can reduce analyses to the essentials they require.
- Users can quickly generate high quality outputs for presentations and journals, without lengthy/repeated formatting.
- Output maps and plots can drive collaborative MPA siting discussions with stakeholders and fisheries managers.

"Prey Mr Babbage, if you put into the machine wrong figures, will right answers come out?"

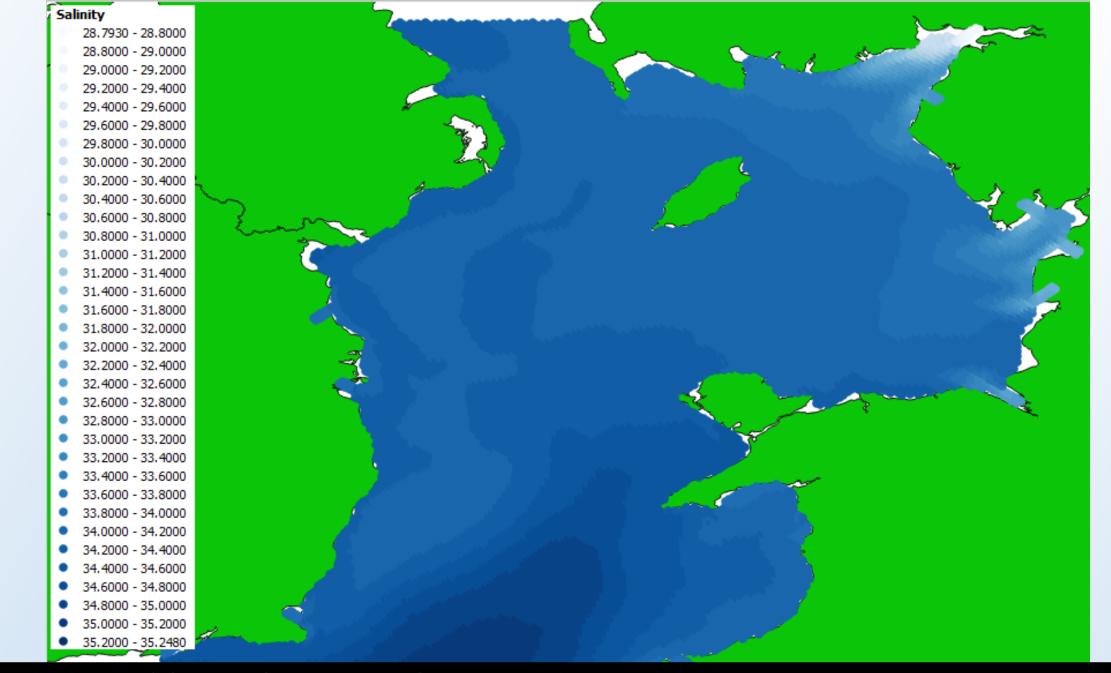




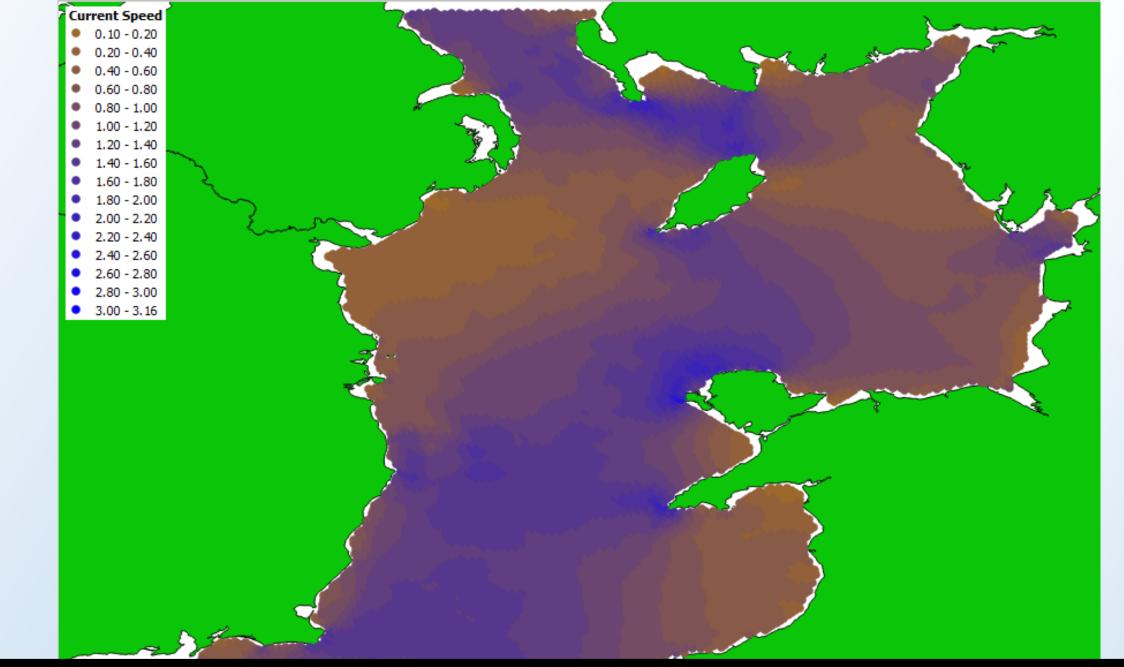
Depth: 391,568 275x455m grids, European Marine Observation and Data Network



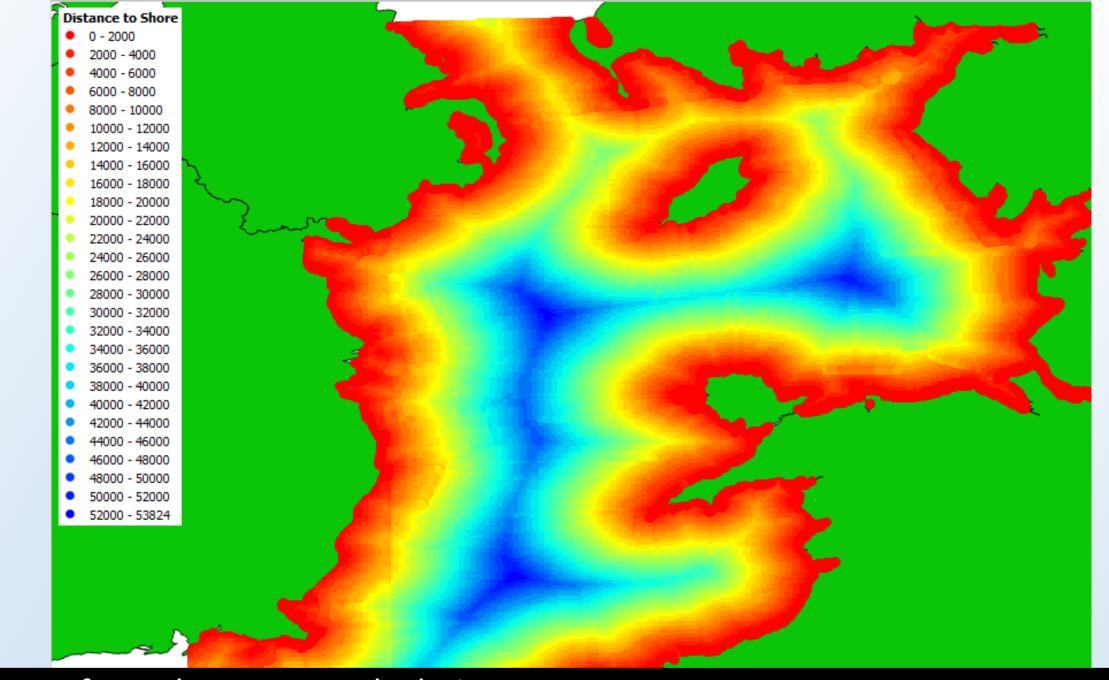
Average monthly sea bottom temperature 2010-2012: 22506 1185x1680m grids, Marine Institute



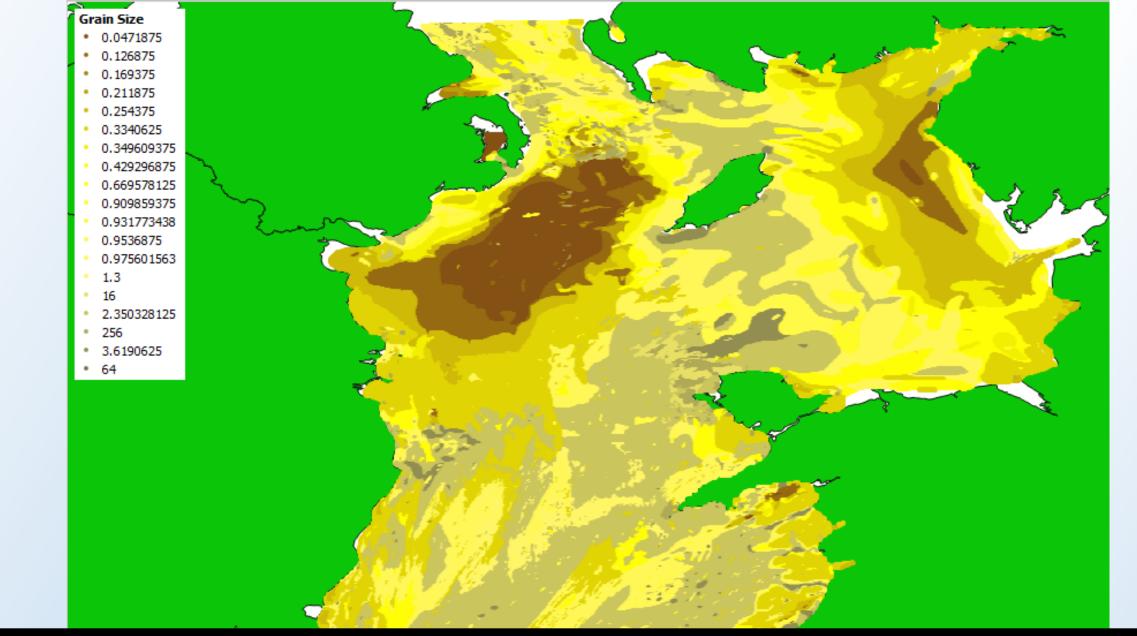
Average monthly sea bottom salinity 2010-2012: 22506 1185x1680m grids, MI



Maximum monthly bottom current speed 2010-2012: 22506 1185x1680m grids, MI



### Distance from shore: map calculation



Grain size: ~250m minimum resolution, British Geological Survey (converted from sediment type classifications)

Thornback ray (*Raja clavata*): 140cm max length Permission to use graphics kindly granted by Marc Dando wildlifeillustrator.com

Blonde ray (*Raja* brachyura): 154cm max length

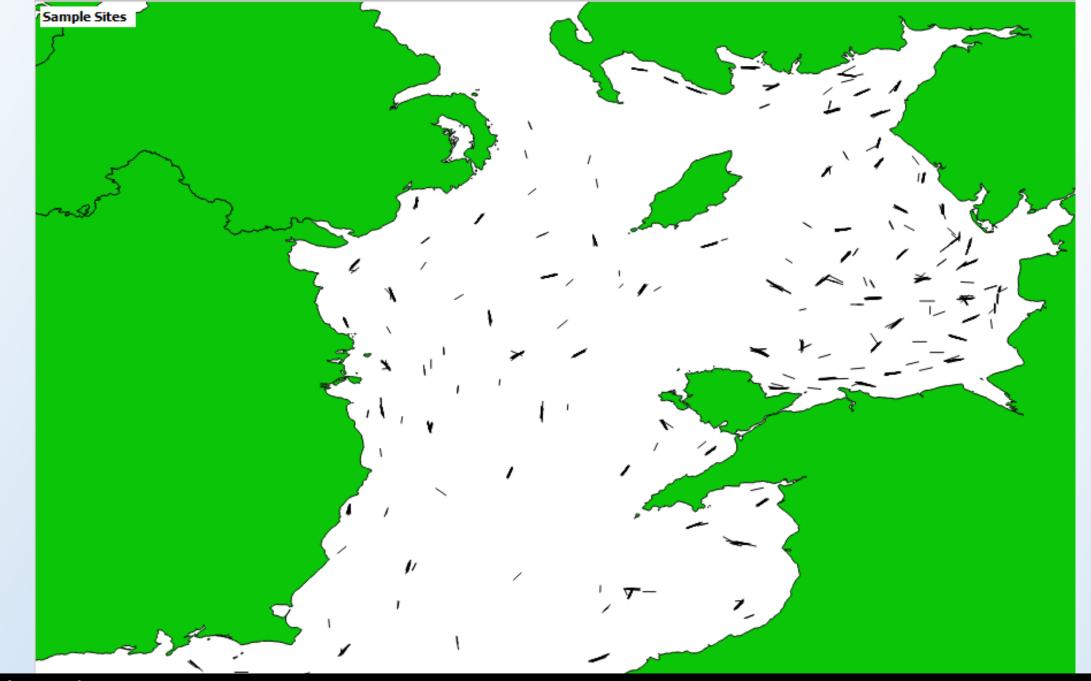
## The study

subject species

Cuckoo ray (Leucoraja naevus): 92cm max length

-

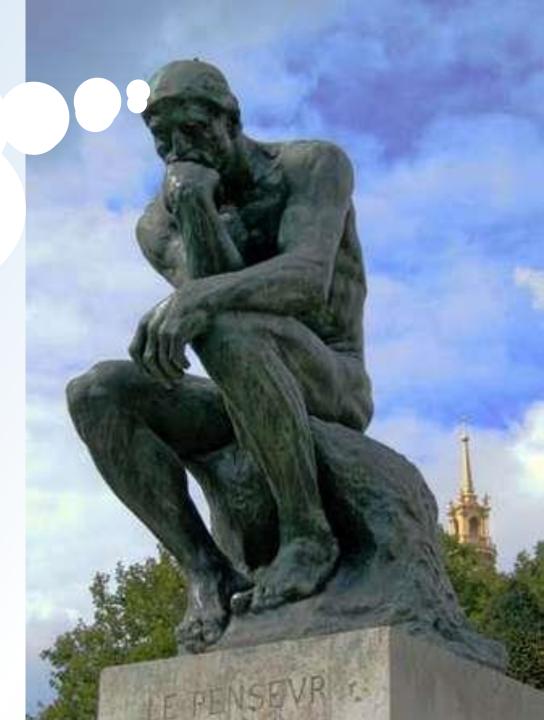
Spotted ray (*Raja montagui*): 78cm max length



Ray abundance at 1447 survey sites: ICES DATRAS, 1993-2012

At what point does making complex stats increasingly available risk a glut of experts testing and improving the models?

People in glass houses shouldn't throw stones... maybe Elith et al would say the same about me!

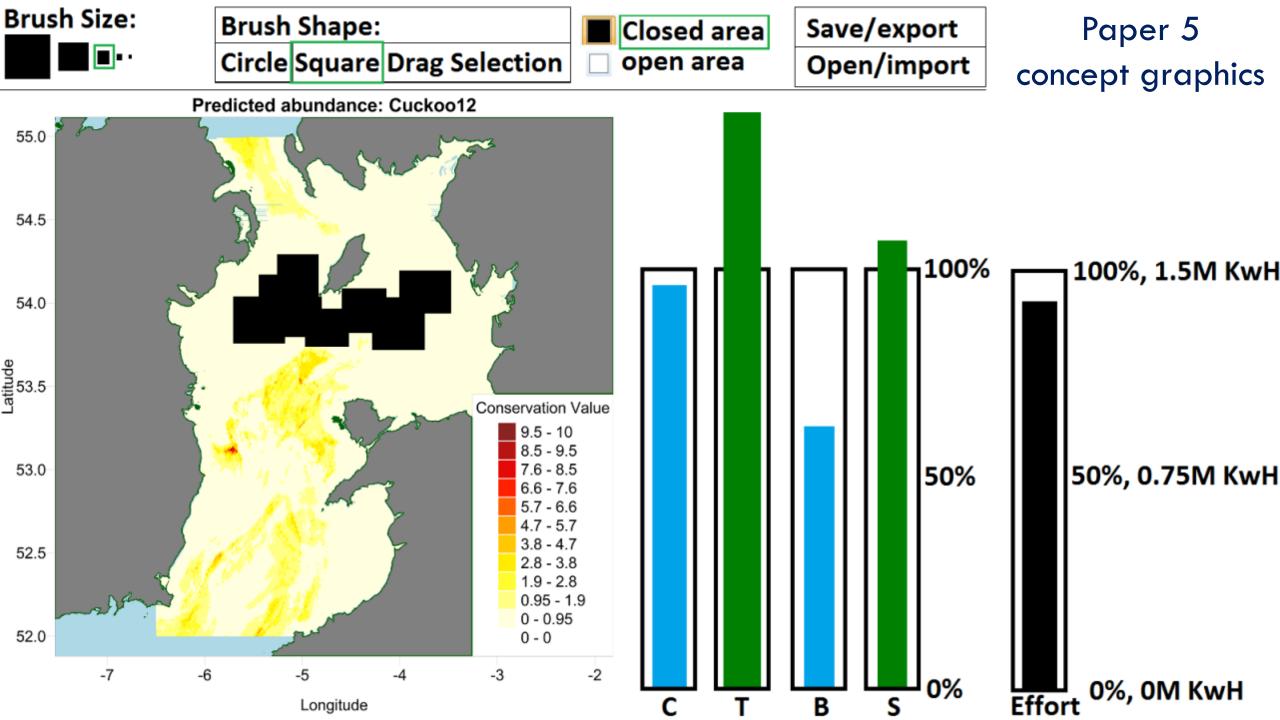


## Novel methods improve prediction of species' distributions from occurrence data

Jane Elith\*, Catherine H. Graham\*, Robert P. Anderson, Miroslav Dudík, Simo@ 22etrier, Antoix Guisan, gdm Robert J. Hijmans, Falk Huettmann, John R. Leathwick, Anthony Lehmann, Jin Li, Lucia G. Lohwann, Bette A. Loiselle, Glenn Manion, Craig Moritz, Miguel Nakamura, Yoshinori Nakazawa, Jacob Mc& Overton, BRT OM-GARP mars-comm A. Townsend Peterson, Steven J. Phillips, Karen Richardson, Ricardo Scachetti-Pereira, Robert E. Schalter, MARS Jorge Soberón, Stephen Williams, Mary S. Wisz and Niklaus E. Zimmermann GLM GDMSS BRUTO 망<sup>0.18</sup> GAN DKGARP Fig. 3. Mean AUC vs mean correlation (COR) for modelling 0.16 methods, summarised across all MARS-INT species. The grey bars are standard BIOCLIN errors estimated in the GLMM (see DOMAII Appendix), reflecting variation for an average species in an average 0.14 LIVES region. The labels are broad classifications of the methods: grey underlined = only use presence data, black capitals = use presence 0.12 and background samples, black 0.65 0.67 0.69 0.71 0.73 0.75 lower case italics = community methods. AUC

### ToDo List: improvements, additions, bugs

- Parallelisation: the core BRT function is a sequential process i.e. single thread only, but could run both halves of a delta model simultaneously.
- OS compatibility
- Swept area AND Spatial error implicit in input data
- Processing time estimate
- Parameter optimisation



### What I'm doing next

- Farallon Institute, Petaluma, CA
- Developing a population dynamics model on forage fish (central northern stock of northern anchovy) abundance in relation to environmental conditions, fisheries exploitation & trophic (predatorprey) interactions in the Southern California Current System using available acoustic & trawl survey data (CalCOFI)
- Explain state shifts
- Non-stationary model, Bayesian TMB? Spatial? Range expansion / contraction
- Sardine eat anchovy eggs...

### Thanks. Any questions?

- Entire project coded in R & requires minimal R knowledge github.com/SimonDedman/gbm.auto
- Code / figures / contact / everything: simondedman.com simondedman@gmail.com
- Ecological Modelling 312 (2015) 77–90: Modelling abundance hotspots for data-poor Irish Sea rays
- Fishes 2 12 (2017)1-22: Advanced spatial modelling to inform management of data-poor juvenile & adult female rays
- ICES Journal of Marine Science 74:2 (2017) 576-587: Towards a flexible Decision Support Tool for MSY-based Marine Protected Area design for skates and rays
- PLoS ONE 12(12): e0188955: Gbm.auto: a software tool to simplify spatial modelling and Marine Protected Area planning
- Bangley et al.: PLoS ONE (in Review): Delineation and Mapping of Coastal Shark Habitat within Pamlico Sound, NC
- Burke et al.: In Prep: Spatial analysis review of BRUVs data
- Grimmel et al.: In Prep: Assessment of Faunal Communities and Ecosystem Interactions within a Shallow-water Lagoon using BRUVs
- Please let me know criticisms/praise/suggestions by email or in person. Thanks!

Thanks to Dr Chuck Bangley, beta tester

Permission to use ray graphics kindly granted by Marc Dando wildlifeillustrator.com . All maps by

SD

Marine Institute myfish





